Solid-state 17O NMR of thymine: a potential new probe to nucleic acid base pairing

Gang Wu,* Shuan Dong and Ramsey Ida

Department of Chemistry, Queen's University, Kingston, Ontario, Canada K7L 3N6. E-mail: gangwu@chem.queensu.ca; Fax: +1-613-533-6669

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We report the first experimental solid-state 17O NMR and theoretical (B3LYP/6-311++G) study of the 17O electricfield-gradient and chemical shielding tensors in a free nucleic acid base, thymine.**

NMR spectroscopy is an important technique for studying structures of biological macromolecules. Most successful NMR applications have been based primarily on observation of spin- $1/2$ nuclei such as ¹H, ¹³C and ¹⁵N. Although oxygen is also an important element in biological molecules, 17O (*S* = 5/2 and natural abundance = 0.037%) NMR studies are far less common.1 To explore the potential of solid-state 17O NMR spectroscopy in studying organic and biological compounds, we recently investigated a number of important oxygen-containing functional groups such as amides,² urea,³ carboxylic acids,⁴ phenols4 and the oxonium ion.5 Oxygen is also common in nucleic acids. Base pairing between nucleic acid molecules often directly involves oxygen atoms, e.g. G:C and A:U base pairing. To our knowledge, solid-state 17O NMR has not been applied to compounds related to nucleic acids.

As a first step, we report solid-state 17O NMR results for a free nucleic acid base, thymine; see Fig. 1. We synthesized $[2^{-17}O]$ thymine and $[4^{-17}O]$ thymine by acid-catalyzed exchange with H_2 ¹⁷O (40.9 atom% ¹⁷O, ISOTEC, Miamisburg, Ohio) from 5-methyl-2-thiouracil and thymine, respectively. Solid-state 17O NMR spectra were recorded on a Bruker Avance-500 spectrometer operating at 67.78 MHz for 17O nuclei. Fig. 2 shows the experimental and simulated 17O magicangle spinning (MAS) NMR spectra of [2-17O]thymine and [4-17O]thymine. Analysis of these spectra yielded the following ¹⁷O NMR parameters: O2, $\delta_{\text{iso}} = 200 \pm 5$ ppm, $C_{\text{Q}} = 6.65 \pm 10^{-10}$ 0.02 MHz, $\eta_{\text{Q}} = 1.00 \pm 0.02$; O4, $\delta_{\text{iso}} = 325 \pm 5$ ppm, $C_{\text{Q}} =$ 8.40 \pm 0.02 MHz, $\eta_{\text{Q}} = 0.10 \pm 0.02$. Following the standard procedure,^{2,6} we were also able to analyze the stationary $17O$ NMR spectra shown in Fig. 2 and obtain the magnitude and

Fig. 1 (A) Chemical structure of thymine. (B) H-bond environment in crystalline thymine.7 Thymine molecules are related by twofold screw axes. Hydrogen atoms are not shown for clarity.

relative orientation of the 17O chemical shift (CS) tensors. The results are summarized in Table 1.

It can be seen from Table 1 that O2 and O4 exhibit drastically different 17O NMR tensors. In particular, the amide-type oxygen, O4, shows a much larger \hat{C}_Q , 8.40 MHz, than the ureatype oxygen, O2, 6.65 MHz. The difference between the 17O CS tensors for O2 and O4 is also striking. The isotropic 17O chemical shifts for O2 and O4 differ by 125 ppm. In addition, the span ($\Omega = \delta_{11} - \delta_{33}$) of the ¹⁷O CS tensor for O4 is more than twice of that for O2. It is also apparent from Table 1 that, whereas the isotropic 17O chemical shifts measured for O4 in the solid and solution states are essentially identical, the corresponding values for O2 differ by approximately 50 ppm! This is clearly a consequence of the strong intermolecular hydrogen-bonding interaction at O2 in crystalline thymine; see Fig. 1.

In order to evaluate quantitatively the influence of intermolecular hydrogen bonding interactions on the 17O EFG and CS tensors in crystalline thymine, we chose to perform quantum chemical calculations using four different models. Model-I is simply an isolated thymine molecule. Model-II consists of two hydrogen-bonded thymine molecules, **1** and **2** as defined in Fig. 1. Model-III also consists of two thymine molecules, **1** and **3**. Model-IV is a trimeric cluster containing **1**, **2** and **3**. The experimental X-ray diffraction structure of thymine7 was used in all the calculations. The positions of the hydrogen atoms were computed using the standard bond lengths and angles. The density functional theory (DFT) calculations were performed on a PC (400 MHz Pentium II processor, 128 MB RAM, 12 GB of

Fig. 2 Experimental (upper) and simulated (lower) 17O MAS NMR spectra of (A) [2-17O]thymine (3753 scans) and (B) [4-17O]thymine (5236 scans). Experimental (upper) and simulated (lower) 17O stationary NMR spectra of (C) [2-17O]thymine (7400 scans) and (D) [4-17O]thymine (5376 scans). The sample spinning frequency was 14.5 kHz. The B_1 field strength at the ¹⁷O frequency was about 70 kHz. Spinning sidebands are marked as 'ssb'. A Hahn-echo pulse sequence was used in acquiring the stationary spectra. The recycle time was 10 s in all experiments.

Table 1 Summary of experimental and theoretical (B3LYP/6-311++G^{**)} ¹⁷O NMR tensors in crystalline thymine

Compound	Model	$\delta_{\rm iso}/\!{\rm ppm}$	δ_{11} /ppm	δ_{22}/ppm	δ_{33} /ppm	C_{Ω} /MHz ^a	$\eta_{\rm Q}$
$[2-17O]$ Thymine		298	441	406	48	8.42	0.55
	П	264	385	359	46	7.82	0.73
	Ш	250	365	344	40	7.72	0.79
	IV	226	327	306	45	7.12	0.99
	Exptl.	200 ± 2 $(247.8)^b$	290 ± 5	270 ± 5	20 ± 5	6.65 ± 0.02	1.00 ± 0.02
$[4-17O]$ Thymine		387	698	487	-25	8.91	0.14
	П	399	685	480	-25	8.83	0.17
	Ш	380	734	496	-33	9.14	0.12
	IV	392	720	491	-34	9.08	0.15
	Exptl.	325 ± 2 $(321.0)^b$	570 ± 5	360 ± 5	20 ± 5	8.40 ± 0.02	0.10 ± 0.02

Fig. 3 Illustration of the orientations of the 17O NMR tensors in thymine.

disk space) using Gaussian 98 program8 with the standard 6-311++G** basis set and the B3LYP exchange functional.9 The theoretical results are also presented in Table 1.

Close examination of the theoretical results reveals a remarkable difference between the 17O NMR tensors for O2 and O4. In particular, both the 17O quadrupole coupling tensor and the CS tensor at O2 exhibit a strong dependence on the cluster model used in the calculation, whereas the 17O NMR tensors at O4 are essentially independent of the model. This clearly reflects the difference in the H-bond environment between O2 and O4. In the discussion that follows, we focus only on the 17O NMR tensors for O2. As seen from Table 1, Model-I predicted Ω = 393 ppm and C_Q = 8.42 MHz for O2, which are considerably larger than the observed values, $\Omega = 270$ ppm and $C_Q = 6.65$ MHz. When two H-bonds were considered in either Model-II or Model-III, smaller values were obtained for Ω and C_O . When a complete H-bond network is included in the calculation (Model-IV), the theoretical results become much closer to the experimental values, $\Omega = 282$ ppm and $C_O = 7.12$ MHz. The observed decrease in the isotropic 17O chemical shift (increase in *shielding*) from Models I to IV results mainly from the changes in δ_{11} and δ_{22} . The large difference between the isotropic 17O chemical shifts measured in the solid state and in solution, 200 *vs*. 247.8 ppm, was well reproduced by the calculations of Model-I and Model-IV. The quadrupole coupling constant exhibits a reduction of approximately 1.3 MHz upon hydrogen bonding, the ¹⁷O EFG tensor increases monotonically from Models I, II, III to IV. Finally, the agreement between the calculated results from Model-IV and the experimental data is reasonable; but it is also clear that all calculated 17O NMR parameters are larger than the observed values by approximately 10%. These discrepancies are likely due to the limitation of the current theory.

Another piece of useful information from the quantum chemical calculations is the absolute orientations of 17O NMR tensors in the molecular frame. As seen from Fig. 3, the *orientations* of the 17O NMR tensors for O2 and O4 are similar, despite the large difference in the magnitude of the individual tensor components. It should be noted that the relative orientation between the 17O EFG and CS tensors shown in Fig. 3 is in agreement with the experimental determination from the analysis of static 17O NMR spectra.

In summary, we have presented the first solid-state 17O NMR study of a free nucleic acid base. The present study demonstrates that it is feasible to obtain solid-state 17O NMR spectra for 17O-labeled nucleobases and that 17O NMR *tensors* are excellent indicators of H-bond formation. These features are potentially useful for probing base pairing in nucleic acids. With the availability of very high magnetic fields (18.8 T or higher) and the advances in solid-state NMR methodology, it is anticipated that solid-state 17O NMR will become a new addition to the arsenal for studying biological macromolecules.

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